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Whole genome association study of brain-wide imaging phenotypes for identifying quantitative trait loci in MCI and AD: A study of the ADNI cohort

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ABSTRACT

A genome-wide, whole brain approach to investigate genetic effects on neuroimaging phenotypes for 30 identifying quantitative trait loci is described. The Alzheimer's Disease Neuroimaging Initiative 1.5 T MRI and 31 genetic dataset was investigated using voxel-based morphometry (VBM) and FreeSurfer parcellation 32 followed by genome-wide association studies (GWAS). One hundred forty-two measures of grey matter 33 (GM) density, volume, and cortical thickness were extracted from baseline scans, GWAS, using PLINK, were 34 performed on each phenotype using quality-controlled genotype and scan data including 530,992 of 620,903 35 single nucleotide polymorphisms (SNPs) and 733 of 818 participants (175 AD, 354 amnestic mild cognitive 36 impairment, MCI, and 204 healthy controls, HC). Hierarchical clustering and heat maps were used to analyze 37 the GWAS results and associations are reported at two significance thresholds ($p < 10^{-7}$ and $p < 10^{-6}$). As 38 expected, SNPs in the APOE and TOMM40 genes were confirmed as markers strongly associated with 39 multiple brain regions. Other top SNPs were proximal to the EPHA4, TP63 and NXPH1 genes. Detailed image 40 analyses of rs6463843 (flanking NXPH1) revealed reduced global and regional GM density across diagnostic 41 groups in TT relative to GG homozygotes. Interaction analysis indicated that AD patients homozygous for the 42 T allele showed differential vulnerability to right hippocampal GM density loss. NXPH1 codes for a protein 43 implicated in promotion of adhesion between dendrites and axons, a key factor in synaptic integrity, the loss 44 of which is a hallmark of AD. A genome-wide, whole brain search strategy has the potential to reveal novel 45candidate genes and loci warranting further investigation and replication. 46

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Introduction

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Data used in the preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (http://www.loni.ucla.edu/ADNI). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. ADNI investigators include (complete listing available at http://www.loni.ucla. edu/ADNI/Collaboration/ADNI_Authorship_list.pdf).

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Recent advances in brain imaging and high throughput genotyping 52techniques enable new approaches to study the influence of genetic 53variation on brain structure and function (Bearden et al., 2007; 54Cannon et al., 2006; Glahn et al., 2007a; Meyer-Lindenberg and 55 Weinberger, 2006; Potkin et al., 2009a). The NIH Alzheimer's Disease 56Neuroimaging Initiative (ADNI) is an ongoing 5-year public-private 57partnership to test whether serial magnetic resonance imaging (MRI), 58positron emission tomography (PET), genetic factors such as single 59

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nucleotide polymorphisms (SNPs), other biological markers, and 60 61 clinical and neuropsychological assessments can be combined to measure the progression of mild cognitive impairment (MCI) and 62 63 early Alzheimer's disease (AD). Given the availability of genomewide SNP data and repeat structural and functional neuroimaging 64 data as part of this initiative, ADNI provides a suitable data set for a 65 large scale imaging genetics study. Using the ADNI baseline MRI data 66 set, we present an imaging genetics framework that employs a whole 67 68 genome and whole brain strategy to systematically evaluate genetic 69 effects on brain imaging phenotypes for discovery of quantitative 70trait loci (QTLs).

Imaging genetics is an emergent transdisciplinary research field 71where the association between genetic variation and imaging 72 73 measures as quantitative traits (QTs) or continuous phenotypes is evaluated. Imaging genetics studies have certain advantages over 74 traditional case control studies. QT association studies have been 75 shown to have increased statistical power and thus decreased sample 76 size requirements (Potkin et al., 2009b). In addition, imaging 77 phenotypes may be closer to the underlying biological etiology of 78 the disease making it easier to identify underlying genes (e.g., Potkin 79 et al., 2009a). Given these observations, the method proposed in this 80 paper focuses on identifying strong associations between regional 81 82 imaging phenotypes as QTs and SNP genotypes as QTLs and aims to provide guidance for refined statistical modeling and follow-up 83 studies of candidate genes or loci. 84

SNPs and other types of polymorphisms in single genes such as 85 APOE have been related to neuroimaging measures in both healthy 86 87 controls and participants with brain disorders such as MCI and AD (e.g., Lind et al., 2006; Wishart et al., 2006). However, the analytic 88 tools that relate a single gene to a few imaging measures are 89 90 insufficient to provide insight into the multiple mechanisms and 91 imaging manifestations of these complex diseases. Genome-wide 92association studies (GWAS) are increasingly performed (Balding, 2006; Hirschhorn and Daly, 2005; Purcell et al., 2007; Zondervan and 93 Cardon, 2007), but effectively relating high throughput SNP data to 94 large scale image data remains a challenging task. As pointed out by 95 96 Glahn et al. (2007b), in imaging genetics, prior studies typically make 97 significant reduction in one or both data types in order to complete analyses. For example, whole brain studies usually focus on a small 98 number of genetic variables (e.g., Ahmad et al., 2006; Brun et al., in 99 press; Filippinia et al., 2009; Nichols and Inkster, 2009; Pezawas et al., 100 101 2004; Shen et al., 2007), while whole genome studies typically examine a limited number of imaging variables (e.g., Baranzini et al., 102 2009; Potkin et al., 2009a; Seshadri et al., 2007). This restriction of 103 target genotypes and/or phenotypes greatly limits our capacity to 104 identify important relationships. 105

106 To overcome this limitation, we present a whole genome and whole brain search strategy for discovering imaging genetics associa-107 tions to guide further detailed analyses. In addition, we present the 108 results from implementation of this technique, including the 109 identification of new genetic loci potentially involved in hippocampal 110 111 and global brain atrophy associated with MCI and AD. In the present 112 study, a detailed set of regions of interest (ROIs) extracted using voxel-based morphometry (VBM) and FreeSurfer automated parcel-113lation defined 142 imaging phenotypes from across the brain 114 (Risacher et al., 2009). A separate GWAS analysis using PLINK 115software (Purcell et al., 2007) was completed for each of these 142 116 imaging phenotypes. Hierarchical clustering and heat maps (Eisen 117 et al., 1998) were used to display and evaluate the association 118 patterns between top SNPs and top imaging phenotypes for multiple 119 statistical thresholds. Subsequent pattern analysis of these heat maps 120not only confirmed prior findings (e.g., APOE and TOMM40 SNPs were 121 among the top ranked list) but also revealed novel QTLs which 122warranted further analyses. Two types of refined imaging genetics 123analysis were performed for one of the top SNPs (NXPH1, rs6463843), 124 125including a VBM analysis assessing global grey matter (GM) density and a regional analysis of target phenotypes. These focused analyses126resulted in interesting imaging genetics findings about the target SNP,127including an overall and regional decrease in GM density associated128with TT genotype relative to the GG genotype with an increased129vulnerability to this effect in AD participants.130

Materials and methods

Sample

Data used in the preparation of this article were obtained from the 133 ADNI database (http://www.loni.ucla.edu/ADNI). The following data 134 from 818 ADNI participants were downloaded from the ADNI 135 database: all baseline 1.5 T MRI scans, the Illumina SNP genotyping 136 data, demographic information, APOE genotype, and baseline diagnosis information. Two participants had genotypic data but no baseline MRI scans and were excluded from all analyses. 139

The ADNI was launched in 2004 by the National Institute on Aging 140(NIA), the National Institute of Biomedical Imaging and Bioengineer-141 ing (NIBIB), the Food and Drug Administration (FDA), private 142 pharmaceutical companies and non-profit organizations, as a \$60 143 million, 5-year public-private partnership. The Principle Investigator 144 of this initiative is Michael W. Weiner, M.D., VA Medical Center and 145 University of California-San Francisco. ADNI is the result of efforts of 146 many co-investigators from a broad range of academic institutions 147 and private corporations. Presently, more than 800 participants, aged 148 55 to 90 years, have been recruited from over 50 sites across the 149United States and Canada, including approximately 200 cognitively 150normal older individuals (i.e., healthy controls or HCs) to be followed 151for 3 years, 400 people with MCI to be followed for 3 years, and 200 152people with early AD to be followed for 2 years. Baseline and 153 longitudinal imaging, including structural MRI scans collected on the 154full sample and PIB and FDG PET imaging on a subset are collected 155every 6-12 months. Additional baseline and longitudinal data 156including other biological measures (i.e. cerebrospinal fluid (CSF) 157markers, APOE and full-genome genotyping via blood sample) and 158clinical assessments including neuropsychological testing and clinical 159examinations are also collected as part of this study. Written informed 160 consent was obtained from all participants and the study was 161 conducted with prior institutional review board's approval. Further 162 information about ADNI can be found in the study of Jack et al. (2008) 163 and Mueller et al. (2005a,b) and at www.adni-info.org. 164

DNA isolation and SNP genotyping

Single nucleotide polymorphism (SNP) genotyping for more than 166 620,000 target SNPs as was completed on all ADNI participants using 167the following protocol. Seven milliliters of blood was taken in EDTA 168containing vacutainer tubes from all participants and genomic DNA 169was extracted using the QIAamp DNA Blood Maxi Kit (Qiagen, Inc., 170Valencia, CA) following the manufacturer's protocol. Lymphoblastoid 171 cell lines were established by transforming B lymphocytes with 172Epstein-Barr virus as described by Neitzel (1986). Genomic DNA 173samples were analyzed on the Human610-Quad BeadChip (Illumina, 174Inc. San Diego, CA) according to the manufacturer's protocols 175(Infinium HD Assay; Super Protocol Guide; Rev. A, May 2008). Before 176initiation of the assay, 50 ng of genomic DNA from each sample was 177 examined qualitatively on a 1% Tris-acetate-EDTA agarose gel to 178 check for degradation. Degraded DNA samples were excluded from 179 further analysis. Samples were quantitated in triplicate with Pico-180 Green® reagent (Invitrogen, Carlsbad, CA) and diluted to 50 ng/µl in 181 Tris-EDTA buffer (10 mM Tris, 1 mM EDTA, pH 8.0). DNA (200 ng) was 182 then denatured, neutralized, and amplified for 22 h at 37 °C (this is 183 termed the MSA1 plate). The MSA1 plate was fragmented with FMS 184 reagent (Illumina) at 37 °C for 1 h, precipitated with 2-propanol, and 185 incubated at 4 °C for 30 min. The resulting blue precipitate was 186

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resuspended in RA1 reagent (Illumina) at 48 °C for 1 h. Samples were 187 then denatured (95 °C for 20 min) and immediately hybridized onto 188 the BeadChips at 48 °C for 20 h. The BeadChips were washed and 189 190subjected to single base extension and staining. Finally, the BeadChips were coated with XC4 reagent (Illumina), dessicated, and imaged on 191 the BeadArray Reader (Illumina). The Illumina BeadStudio 3.2 192software was used to generate SNP genotypes from bead intensity 193data. All SNP genotypes are publicly available for download at the 194195ADNI website (http://www.loni.ucla.edu/ADNI).

MRI analysis and extraction of imaging phenotypes 196

Two widely employed automated MRI analysis techniques were 197198 used to process and extract brain-wide target MRI imaging phenotypes from all baseline scans of ADNI participants as previously 199described (Risacher et al., 2009). First, voxel-based morphometry 200 (VBM; Ashburner and Friston, 2000; Good et al., 2001; Mechelli et al., 2012005) was performed to define global grey matter (GM) density maps 202 and extract local GM density values for 86 target regions (Table 1). 203 Second, automated parcellation via FreeSurfer V4 (http://surfer.nmr. 204 mgh.harvard.edu/) was conducted to define 56 volumetric and 205cortical thickness values (Table 2). All included ADNI participants 206 had a minimum of two 1.5 T MP-RAGE scans at baseline following the 207ADNI MRI protocol (Jack et al., 2008). Each raw scan was indepen-208 dently processed using FreeSurfer and VBM. 209

210 For VBM analysis, SPM5 (http://www.fil.ion.ucl.ac.uk/spm/) was 211 used to create an unmodulated normalized GM density map $(1 \times 1 \times 1 \text{ mm voxel size, } 10 \text{ mm FWHM Gaussian kernel for})$ 212 smoothing) in the MNI space for each scan as previously described 213(Risacher et al., 2009). A mean GM density map was created as an 214

average of two independent smoothed, unmodulated normalized GM 215 density maps for each participant using SPM5. The MarsBaR region of 216 interest (ROI) toolbox (Brett et al., 2002; Tzourio-Mazover et al., 217 2002) as implemented in SPM5 was then used to extract a single 218 mean GM density value for 86 target regions in MNI space (Table 1) to 219be used as target QTs for the imaging genetic analyses. In addition to 220 the individual MarsBaR ROIs, larger target regions defined by 221 combining the mean GM density value from a set of MarsBaR ROIs 222 were used as imaging phenotypes. All individual and combined mean 223 GM density values are referred to as VBM phenotypes; see Table 1 for a 224 total list and explanation of the 86 VBM phenotypes. 225

For automated segmentation and parcellation, FreeSurfer V4 was 226 employed to automatically label cortical and subcortical tissue classes 227using an atlas-based Bayesian segmentation procedure (Dale et al., 228 1999; Fischl and Dale, 2000; Fischl et al., 2002, 1999) and to extract 229 target region volume and cortical thickness, as well as to extract total 230 intracranial volume (ICV) for all participants. Extracted FreeSurfer 231 values for two independently processed MP-RAGE images of the same 232participant were averaged to create a mean value for volumetric and 233cortical thickness measures for all target regions. Mean volumetric 234and cortical thickness measures extracted using automated parcella-235tion are referred to as *FreeSurfer phenotypes*; see Table 2 for a total list 236 of the 56 FreeSurfer phenotypes defined for selected target regions. 237

Genome-wide association analysis of imaging phenotypes

APOE genotype

The APOE gene is an important target gene in AD research (Farrer 240 et al., 1997). However, the two previously identified APOE SNPs 241 important in AD susceptibility (rs429358, rs7412) were not available 242

t1.1 Table 1

VBM phenotypes defined as mean GM densities of various regions of interest (ROIs). SPM5 was applied for computing voxel-wise GM density values, while the MarsBaR ROI toolbox was used to define ROIs in the MNI space. A total number of 43×2 = 86 phenotypes were calculated. Each of the 43 IDs shown in the table corresponds to two phenotypes: one for the left side and the other for the right side. For example, "LAmygdala" indicates the mean GM density of the left amygdala. Each region marked with * in the table is a combined set of more than one MarsBaR ROI. For example, "RMeanLatTemporal" indicates the mean GM density of the right lateral temporal region defined by a set of MarsBaR ROIs, including right inferior temporal gyrus, right middle temporal gyrus, and right superior temporal gyrus.

t1.2 t1.3	Phenotype ID	Region of interest (Phenotype is defined as the mean GM density of the ROI)	Phenotype ID	Region of interest (Phenotype phenotype is defined as the mean GM density of the ROI)				
t1.4	Amygdala	Amygdala	MidTempPole	Middle temporal pole				
t1.5	Angular	Angular gyrus	MidTemporal	Middle temporal gyrus				
t1.6	AntCingulate	Anterior cingulate	Olfactory	Olfactory gyrus				
t1.7	Fusiform	Fusiform gyrus	Parahipp	Parahippocampal gyrus				
t1.8	Heschl	Heschl's gyrus	PostCingulate	Posterior cingulate				
t1.9	Hippocampus	Hippocampus	Postcentral	Postcentral gyrus				
t1.10	InfFrontal_Oper	Inferior frontal operculum	Precentral	Precentral gyrus				
t1.11	InfFrontal_Triang	Inferior frontal triangularis	Precuneus	Precuneus				
t1.12	InfOrbFrontal	Inferior orbital frontal gyrus	Rectus	Rectus gyrus				
t1.13	InfParietal	Inferior parietal gyrus	Rolandic_Oper	Rolandic operculum				
t1.14	InfTemporal	Inferior temporal gyrus	Supfrontal	Superior frontal gyrus				
t1.15	Insula	Insula	SupOrbfrontal	Superior orbital frontal gyrus				
t1.16	Lingual	Lingual gyrus	SupParietal	Superior parietal gyrus				
t1.17	MedOrbFrontal	Medial orbital frontal gyrus	SupTempPole	Superior temporal pole				
t1.18	MedSupFrontal	Medial superior frontal gyrus	SupTemporal	Superior temporal gyrus				
t1.19	MidCingulate	Middle cingulate	SuppMotorArea	Supplementary motor area				
t1.20	MidFrontal	Middle frontal gyrus	Supramarg	Supramarginal gyrus				
t1.21	MidOrbFrontal	Middle orbital frontal gyrus	Thalamus	Thalamus				
t1.22								
t1.23	Phenotype ID	Regions of interest (phenotype is defined as the av	verage GM density of multiple M	IarsBaR ROIs)				
t1.24	MeanCing*	Anterior cingulate, middle cingulate, and posterior	cingulate					
t1.25	MeanFrontal*	Inferior frontal operculum, inferior orbital frontal	gyrus, inferior frontal triangulari	s, medial orbital frontal gyrus, middle frontal gyrus,				
	middle orbital frontal gyrus, superior frontal gyrus, medial superior frontal gyrus, superior orbital frontal gyrus, rectus gyrus, r							
		operculum, and supplementary motor area						
t1.26	MeanLatTemporal*	Inferior temporal gyrus, middle temporal gyrus, ar	Inferior temporal gyrus, middle temporal gyrus, and superior temporal gyrus					
t1.27	MeanMedTemporal*	Amygdala, fusiform gyrus, Heschl's gyrus, hippocan	npus, lingual gyrus, olfactory gyru	us, parahippocampal gyrus, middle temporal pole, and				
		superior temporal pole						
t1.28	MeanOccipital*	Calcarine gyrus, cuneus, inferior occipital gyrus, m	Calcarine gyrus, cuneus, inferior occipital gyrus, middle occipital gyrus, and superior occipital gyrus					
t1.29	MeanParietal*	Angular gyrus, inferior parietal gyrus, superior par	ietal gyrus, supramarginal gyrus	, and precuneus				
t1.30	MeanTemporal*	Amygdala, fusiform gyrus, Heschl's gyrus, hippoca middle temporal gyrus, middle temporal pole, sup	Amygdala, fusiform gyrus, Heschl's gyrus, hippocampus, lingual gyrus, olfactory gyrus, parahippocampal gyrus, inferior temporal gyrus, middle temporal gyrus, middle temporal pole, superior temporal pole, and superior temporal gyrus					

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t2.1 Table 2

FreeSurfer phenotypes defined as volumetric or cortical thickness measures of various regions of interest (ROIs). FreeSurfer was applied for automated parcellation to extract volume and cortical thickness values for a total number of $28 \times 2 = 56$ ROIs. Each of the 28 IDs shown in the table corresponds to two phenotypes: one for the left side and the other for the right side. For example, "LAmygVol" indicates the volume of the left superior temporal gyrus.

t2.2 t2.3	Phenotype ID	Phenotype description
t2.4	AmygVol	Volume of amygdala
t2.5	CerebCtx	Volume of cerebral cortex
t2.6	CerebWM	Volume of cerebral white matter
t2.7	HippVol	Volume of hippocampus
t2.8	InfLatVent	Volume of inferior lateral ventricle
t2.9	LatVent	Volume of lateral ventricle
t2.10	EntCtx	Thickness of entorhinal cortex
t2.11	Fusiform	Thickness of fusiform gyrus
t2.12	InfParietal	Thickness of inferior parietal gyrus
t2.13	InfTemporal	Thickness of inferior temporal gyrus
t2.14	MidTemporal	Thickness of middle temporal gyrus
t2.15	Parahipp	Thickness of parahippocampal gyrus
t2.16	PostCing	Thickness of posterior cingulate
t2.17	Postcentral	Thickness of postcentral gyrus
t2.18	Precentral	Thickness of precentral gyurs
t2.19	Precuneus	Thickness of precuneus
t2.20	SupFrontal	Thickness of superior frontal gyrus
t2.21	SupParietal	Thickness of superior parietal gyurs
t2.22	SupTemporal	Thickness of superior temporal gyrus
t2.23	Supramarg	Thickness of supramarginal gyrus
t2.24	TemporalPole	Thickness of temporal pole
t2.25	MeanCing	Mean thickness of caudal anterior cingulate,
		isthmus cingulate, posterior cingulate, and
		rostral anterior cingulate
t2.26	MeanFront	Mean thickness of caudal midfrontal, rostral
		midfrontal, superior frontal, lateral orbitofrontal,
		and medial orbitofrontal gyri and frontal pole
t2.27	MeanLatTemp	Mean thickness of inferior temporal, middle temporal,
	· · · · ·	and superior temporal gyri
t2.28	MeanMedTemp	Mean thickness of fusiform, parahippocampal,
		and lingual gyri, temporal pole and transverse
		temporal pole
t2.29	MeanPar	Mean thickness of inferior and superior parietal gyri,
		supramarginal gyrus, and precuneus
t2.30	MeanSensMotor	Mean thickness of precentral and postcentral gyri
t2.31	MeanTemp	Mean thickness of interior temporal, middle temporal,
		superior temporal, fusiform, paranippocampal, and
		lingual gyri, temporal pole and transverse temporal pole

on the Illumina array. Therefore, we determined the genotypes of the two APOE SNPs (rs429358, rs7412) using the APOE $\epsilon 2/\epsilon 3/\epsilon 4$ status information from the ADNI clinical database for each participant.

246 Quality control

The original genotype data contained 620,903 markers, including 247620,901 genomic markers on the Illumina chip plus 2 APOE SNPs 248whose values were obtained from the APOE status data. Only SNP 249markers were analyzed in this study. The following quality control 250251(QC) steps were performed on these genotype data using the PLINK 252software package (http://pngu.mgh.harvard.edu/~purcell/plink/), release v1.06. SNPs were excluded from the imaging genetics 253analysis if they could not meet any of the following criteria: (1) 254call rate per SNP \geq 90%, (2) minor allele frequency (MAF) \geq 5%, and 255(3) Hardy–Weinberg equilibrium test of $p \le 10^{-6}$ using healthy 256control (HC) subjects only. Participants were excluded from the 257analysis if any of the following criteria was not satisfied: (1) call rate 258per participant \geq 90% (1 participant was excluded); (2) gender 259check (2 participants were excluded); and (3) identity check (3 260sibling pairs were identified with PI_HAT over 0.5; one participant 261 from each pair was randomly selected and excluded). Population 262stratification analysis suggested the advisability of restricting 263analyses to non-Hispanic Caucasians (79 participants were excluded 264 265 from this report). After the QC procedure, 733 out of 818 participants and 530,992 out of 620,903 markers remained in the analysis and the overall genotyping rate for the remaining dataset was over 99.5%. 268

GWAS analyses

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One hundred forty-two separate GWAS analyses on 142 selected 270 imaging phenotypes (86 VBM phenotypes and 56 FreeSurfer 271phenotypes) were completed using the quality-controlled SNP data. 272 All the imaging phenotypes were adjusted for the baseline age, 273 gender, education, handedness, and baseline intracranial volume 274(ICV) using the regression weights derived from the HC participants. 275prior to any of the GWAS analyses (Risacher et al., 2009). Using the 276PLINK software package (v1.06) with the quantitative trait association 277option, each GWAS analysis calculated the main effects of all SNPs on 278the target quantitative imaging phenotype. An additive SNP effect was 279assumed and the empirical *p*-values were based on the Wald statistic 280 (Purcell et al., 2007). Right hippocampal GM density was selected for a 281 detailed sample analysis of a target QC because it had the largest 282 number of associations at $p < 10^{-6}$. A Manhattan plot and a quantile-283 quantile (Q–Q) plot were used to visualize GWAS results for the right 284hippocampal GM density. All association results surviving the 285significance threshold of $p < 10^{-6}$ were saved and prepared for 286additional pattern analysis. 287

Sample definition and demographics

The sample employed in the GWAS analyses of FreeSurfer 280 phenotypes included participants that passed the genotype QC 290procedure and FreeSurfer processing. The sample used in the GWAS 291 analyses of VBM phenotypes included participants that passed the 292genotype QC procedure, FreeSurfer processing, and VBM processing. 293Demographic information, including baseline age, years of education, 294gender distribution, and handedness distribution, was compared 295between baseline diagnostic groups for each sample separately using 296one-way ANOVAs and chi-squared analyses as applicable in SPSS 297(version 16.0.1). 298

Pattern analyses of GWAS results

and data reduction for

To expedite the review of GWAS results and data reduction for 300 subsequent analyses, we employed heat map and hierarchical 301 clustering approaches (Eisen et al., 1998; Levenstien et al., 2003; 302 Sloan et al., submitted for publication) for visualizing associations 303 between identified SNPs and their associated imaging phenotypes 304 at various significance levels. Heat maps are colored images 305 mapping given values (in this study, $-\log_{10}(p)$ of the 306 corresponding association) to coded colors. Generally, heat maps 307 have dendrograms, representing hierarchical clustering results 308 along both the x-axis and y-axis (in this study, x: imaging 309 phenotypes, y: SNPs). R (v.2.9.0) (http://www.r-project.org/), an 310 open source statistical computing package, was employed to create 311 the heat maps. Hierarchical clustering was completed using Eucli-312 dean distance methods to define dissimilarity between two nodes 313 and average of distances between all pairs of objects in two clusters to 314 measure the distance between two clusters. On each heat map, 315 significant associations between imaging phenotypes and SNPs 316 were marked with an "x" to facilitate visual evaluation of the 317 results. The color bar on the left side of the heat map encodes the 318 chromosome IDs for the corresponding SNPs. In addition to the 319 heat maps, a summary statistic detailing the number of significant 320 associations at the $p < 10^{-6}$ level for each imaging phenotype and 321 SNP was evaluated to help guide the refined analyses. In the 322 present study, all imaging GWAS results are presented and 323 analyzed using heat maps and summary statistics. 324

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325 Detailed analysis of a target SNP identified by cluster analysis

An in-depth analysis was performed for one of the top SNPs selected by inspecting the heat maps and summary statistics. The refined analysis included two steps: (1) a global voxel-based analysis on the entire brain using VBM and (2) regional analyses of identified target phenotypes. We included both types of analyses as they provide complementary information relevant to assessing risk for AD or disease progression (Risacher et al., 2009; Saykin et al., 2006).

For global analyses, VBM was performed on a voxel-by-voxel basis 333 334using a general linear model (GLM) approach as implemented in SPM5. After identifying the SNP of interest, a two-way ANOVA assessing the 335effects of baseline diagnostic group and SNP genotype value was 336 337 performed to compare the smoothed, unmodulated normalized GM maps to determine any significant effects of diagnosis, SNP genotype, 338 and SNP-by-diagnosis interactions on global GM density between and 339 within groups. Contrasts between genotypes were displayed with a 340 significance threshold of p < 0.01 corrected for multiple comparisons 341 using a false discovery rate (FDR) technique when including the entire 342 sample. For contrasts within a single diagnostic group, the p < 0.01 (FDR) 343 threshold was too stringent given the reduced power and no significant 344 voxels were observed. Therefore, we used a slightly less stringent 345 346 significance threshold of p < 0.001 (uncorrected for multiple comparisons) when examining SNP effects within a diagnostic group, in order to 347 evaluate the pattern of GM density associated with genotype. A 348 minimum cluster size (k) of 27 voxels was required for significance in 349all comparisons and an explicit GM mask was used to restrict analyses to 350 351GM regions. Age, gender, education, handedness and baseline ICV were included as covariates in all analyses. 352

For ROI analyses, a two-way multivariate ANOVA in SPSS (version 353 16.0.1) was completed to determine the effect of baseline diagnosis 354355 and genotype on bilateral hippocampal and mean medial temporal 356lobar GM density. Similar to the VBM analysis, age, gender, education, handedness, and baseline ICV were included as covariates in all 357 comparisons. Independent effects of baseline diagnosis and genotype, 358 as well as the interaction effect of baseline diagnosis × genotype for 359 each SNP, were assessed for selected imaging variables. All graphs 360 were created using SigmaPlot (version 10.0). 361

362 Results

363 Sample characteristics after QC

After quality control of the genotyping data including the 364 exclusion of 79 participants to avoid potential population stratifica-365 tion confounds, 733 out of 818 ADNI participants remained in the 366 present study. Among these 733 participants, 729 sets of scans were 367 successful in FreeSurfer segmentation and parcellation and were 368 included in GWAS analyses of FreeSurfer phenotypes (56 volumetric 369 and cortical thickness values described in Table 2). Seven hundred 370 fifteen participants had successful VBM processing and were used in 371 372 GWAS analyses of VBM phenotypes (86 GM density values described 373 in Table 1). Table 3 shows the demographics information of the sample analyzed for both FreeSurfer and VBM studies. In both 374 samples, gender and education are significantly different (overall 375 p < 0.05) among baseline diagnostic groups (HC, MCI, AD). In the subsequent GWAS analyses, baseline age and gender, as well as education, handedness, and baseline ICV are included as covariates. 378

GWAS of imaging phenotypes

For convenience, in this paper, an SNP is described by its rs number 380 together with its respective gene (i.e., the closest gene, as annotated in 381 Illumina's Human610-Quad SNP list). Shown in Fig. 1 are all the 382 imaging genetics associations at a significance threshold of $p < 10^{-7}$ (a 383 typical threshold for genome-wide significance), which are discovered by GWAS analysis of 142 imaging phenotypes (i.e., quantitative 385 traits, or QTs).

At the $p < 10^{-7}$ significance level, 22 strong SNP-QT associations 387 (see blocks labeled with "x" in Fig. 1) were identified in the GWAS 388 analyses, and five SNPs were involved in these associations. As a well-389 established AD risk factor (Farrer et al., 1997), the APOE SNP rs429358 390 confirmed to have multiple associations with both FreeSurfer QTs and 391 VBM QTs, showing as the most prominent imaging genetics pattern at 392 the significance level of $p < 10^{-7}$. In addition, associations with 393 multiple FreeSurfer QTs were identified for rs2075650 (TOMM40), 394 supporting the recent finding of TOMM40 as a gene adjacent to APOE 395 and an additional contributor to AD (Osherovich, 2009; Potkin et al., 396 2009a). Three additional SNPs were found to have strong associations 397 with one or more VBM QTs: rs6463843 (NXPH1), rs4692256 398 (LOC391642), and rs10932886 (EPHA4). Further information about 399 these SNPs is available in Table 4. 400

A number of imaging phenotypes were identified to have strong 401 associations with target SNPs in the GWAS analyses, suggesting that 402 these values may be sensitive QTs to imaging genetics studies of AD. As 403 expected, both the left and right amygdalar and hippocampal regions 404 were found to be strongly associated with rs429358 (APOE) using 405 volumetric and GM density measures. In addition, rs2075650 406 (TOMM40) was significantly associated with bilateral hippocampal 407 volume and left amygdalar volume. Additional imaging phenotypes 408 found to be sensitive QTs, include (a) volume measures from the right 409 cerebral cortex and cerebral white matter, (b) cortical thickness mea-410 sures from left and right inferior parietal gyri, and right middle tem-411 poral gyrus, and (c) GM density measures from the left middle orbital 412 frontal gyrus, left precuneus, left superior frontal gyrus, and left and 413 right mean frontal lobe regions (see MeanFrontal definition in Table 1). 414

Heat maps of clustered associations at a somewhat less stringent 415 significance level $(p < 10^{-6})$ are shown in Fig. 2. As expected, more 416 SNPs and OTs are involved. The top 10 SNPs and their respective genes 417 ranked by the total number of significant QT associations at $p < 10^{-6}$ 418 are shown in Table 4. With more SNPs and QTs available in the heat 419 maps, interesting clustering patterns in both the imaging and genetics 420 dimensions were revealed by examining the corresponding dendro-421 grams (i.e., hierarchical clustering results). In the imaging dimension 422 (x-axis), many pairs of left and right measures of the same structure 423 were clustered together, supporting the symmetric relationship 424

t3.1 Table 3

Demographic information and total number of participants involved in each analysis. Of 818 ADNI participants, 733 remained after quality control of the genotyping data and consideration of population stratification. Among these 733 participants, 729 subjects succeeded in FreeSurfer segmentation and parcellation and were involved in the GWAS analysis of FreeSurfer phenotypes. Of these, 715 subjects had successful VBM processing and were involved in the GWAS analysis of VBM phenotypes. Basic demographics information is shown for both groups of participants.

t3.2 t3.3	Category	FreeSurfer pho	FreeSurfer phenotypes (729 subjects)				VBM phenotypes (715 subjects)			
t3.4		НС	MCI	AD	p-value	HC	MCI	AD	p-value	
t3.5	Number of subjects	203	351	175	-	203	346	166	-	
t3.6	Gender (M/F)	111/92	229/122	97/78	0.019	111/92	225/121	90/76	0.017	
t3.7	Baseline age (years; mean \pm SD)	76.1 ± 5.0	75.1 ± 7.3	75.5 ± 7.6	0.283	76.1 ± 5.0	75.1 ± 7.4	75.5 ± 7.6	0.285	
t3.8	Education (years; mean \pm SD)	16.1 ± 2.7	15.7 ± 3.0	14.9 ± 3.0	0.0004	16.1 ± 2.7	15.7 ± 3.0	14.9 ± 3.0	0.0003	
t3.9	Handedness (R/L)	188/15	318/33	163/12	0.53	188/15	314/32	157/9	0.31	

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Fig. 1. Heat maps of SNP associations with quantitative traits (QTs) at the significance level of $p < 10^{-7}$. GWAS results at a statistical threshold of $p < 10^{-7}$ using QTs derived from FreeSurfer (top) and VBM/MarSBaR (bottom) are shown. $-\log_{10}(p$ -values) from each GWAS are color-mapped and displayed in the heat maps. Heat map blocks labeled with "x" reach the significance level of $p < 10^{-7}$. Only top SNPs and QTs are included in the heat maps, and so each row (SNP) and column (QT) has at least one "x" block. Dendrograms derived from hierarchical clustering are plotted for both SNPs and QTs. The color bar on the left side of the heat map codes the chromosome IDs for the corresponding SNPs. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

between these phenotypes and genetic variation. In addition, regional
similarity was also detected including a prominent pattern of multiple
orbital frontal measures clustered together in Fig. 2b. In the genomic
dimension (*y*-axis), three SNPs from LOC391642 were grouped
together in Fig. 2b, suggesting an increased likelihood of linkage
disequilibrium (LD) effects.

431 Refined analysis for a sample target QT

Subsequent analyses focused on a target QT and a target SNP
selected from heat maps in Fig. 2. Shown in Fig. 3 are the Manhattan
and Q-Q plots of the GWAS for the target QT, right hippocampal GM

density (RHippocampus in Fig. 2b). In the Q–Q plot, for most of the p-435values, the observed p-values from GWAS are almost the same as the436expected p-values from the null hypothesis. There was little or no437evidence of systematic bias, which could be caused by factors such as a438strong population substructure and genotyping artifacts. The p-values439in the upper tail of the distribution do show a significant deviation440suggesting strong associations between these SNPs and the QT.441

Refined analysis for a sample target SNP

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A target SNP, rs6463843 (NXPH1), was selected for detailed 443 imaging analyses since it was the only SNP strongly associated with 444

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t4.1 Table 4

Top quantitative trait (QT) loci ranked by the total number of associations at the significance level of $p < 10^{-6}$. Relevant information about top ranked SNPs and their respective genes (i.e., the closet gene, as annotated in Illumina's Human610-Quad SNP list (except APOE information extracted from dbSNP)) is shown in this table, including SNP, chromosome (CHR), coordinate (Build 36.2), gene, location, and position. In addition, the number of QTs that are associated with each SNP at the significance level of $p < 10^{-6}$ is also shown. The SNPs are ordered according to the last column.

t4.2 t4.3	SNP	CHR	Coordinate	Gene	Location	Position	Number o	Number of QT associations		
t4.4							VBM	FreeSurfer	Total	
t4.5	rs10932886	2	221428332	EPHA4	Flanking_3UTR	- 562,659	27	0	27	
t4.6	rs429358	19	50103781	APOE	Coding	Exon 4	4	15	19	
t4.7	rs7610017	3	190826118	TP63	Flanking_5UTR	-5792	19	0	19	
t4.8	rs6463843	7	8805242	NXPH1	Flanking_3UTR	-46124	9	0	9	
t4.9	rs2075650	19	50087459	TOMM40	Intron	-31	0	5	5	
t4.10	rs16912145	10	59752674	UBE2D1	Flanking_5UTR	-12071	4	0	4	
t4.11	rs12531488	7	144523019	LOC643308	Flanking_5UTR	-154052	3	0	3	
t4.12	rs7526034	1	63359561	LOC199897	Flanking_5UTR	- 103696	0	2	2	
t4.13	rs7647307	3	69705878	LOC642487	Flanking_5UTR	-31337	0	2	2	
t4.14	rs4692256	4	27353816	LOC391642	Flanking_3UTR	-156945	1	0	1	

both left and right hippocampi other than rs429358 (APOE) and 445 rs2075650 (TOMM40). The results of a two-way ANOVA using VBM to 446 compare the effects of baseline diagnostics group and rs6463843 447 (NXPH1) genotype on global GM density are shown in Fig. 4. After 448 449 evaluating hippocampal GM density group means for each diagnosisgenotype group, we chose to contrast GG vs. TT (GG>TT) using all 450participants (*n* = 715; 166 AD (44 TT, 78 GT, 44 GG); 346 MCI (82 TT, 451170 GT, 94 GG); 203 HC (35 TT, 105 GT, 63 GG)). As shown in Fig. 4a, 452TT participants had significantly reduced global GM density through-453454out the brain relative to GG participants (p < 0.01 (FDR), k = 27). Maximal differences between groups were found in a number of 455regions known to be associated with AD, including the medial 456temporal lobe (-36, -30, -17; T=5.20) and frontal (19, 56, 457-15; T = 5.56), parietal (26, -59, 67; T = 5.71) and temporal (-59, 4584592, -30; T=4.81) lobe cortical surfaces. In order to determine whether a particular diagnostic group was responsible for the effects 460 seen in the full sample contrast of GG>TT, we evaluated the same 461 comparison within each baseline diagnostic group (Fig. 4b; AD, MCI, 462 HC). The pattern of significant voxels for GG>TT was largest in the 463 464AD group, with highly significant clusters in the right hippocampus (31, -26, -15; T=5.34), left medial temporal lobe (-25, -32, -32, -32)465 -7; T = 4.37), and frontal lobe (-35, 49, -13; T = 4.33). MCI and 466 HC groups also showed significant voxels in the contrast of GG>TT, 467 468 with maximum voxels found in the inferior frontal lobe (45, 25, -13;T=3.82) and middle frontal lobe (-25, 6, 62; T=4.58), respectively. 469 470 The AD panel in Fig. 4b showed more prominent patterns, while the 471 MCI and HC panels appeared less structured. This suggested a possible SNP-by-diagnosis interaction effect on brain structure, 472 473 which is examined below at a more detailed level for several candidate imaging phenotypes. Furthermore, the inclusion of APOE 474 genotype as a covariate did not significantly alter these effects (data 475not shown). 476

Based on the heat map and VBM results, four GM density 477 478 measures were further evaluated as phenotypes for additional associations with rs6463843 (NXPH1). As shown in Fig. 5, expected 479baseline diagnostic differences in left (Fig. 5a; F(7,708) = 79.4, 480 *p*<0.001) and right (Fig. 5b; *F*(7,708) = 78.4, *p*<0.001) hippocampal 481 GM density, as well as left (Fig. 5c; F(7,708) = 60.3, p < 0.001) and 482 483 right (Fig. 5d; F(7,708) = 59.4, p < 0.001) mean medial temporal lobe GM density were found. Pairwise comparisons indicated that AD 484 participants had significantly reduced hippocampal and mean 485 medial temporal lobe GM density relative to both MCI and HC 486 participants (all p < 0.001). MCI participants also showed a signifi-487 cantly reduced GM density in all these regions relative to HCs 488 (p < 0.001). The main effect of genotype across all participants was 489also significant for left and right hippocampal GM density (left, F 490 491 (7,708) = 10.4; right, F(7,708) = 9.9, both p < 0.001) and left and 492 right mean medial temporal lobe GM density (left, F(7,708) = 7.9; right, F(7,708) = 9.0, both p < 0.001). Paired comparisons indicated 493 significantly reduced left and right hippocampal and mean medial 494 temporal lobar GM density in participants with a TT genotype 495 relative to those with a GG genotype in the rs6463843 (NXPH1) SNP 496 (p < 0.01). In addition, participants with the TT genotype had 497 significantly reduced left and right mean medial temporal lobe GM 498 density relative to TG heterozygotes (p < 0.01). The interaction effect 499of baseline diagnosis and rs6463843 genotype was also significant 500 for right hippocampal GM density (p < 0.05), but not for the other 501three regions, which suggested that AD patients with TT genotype 502were particularly vulnerable to increased GM density loss in right 503hippocampus. 504

Discussion

Methodological overview

Employing a whole genome and entire brain strategy, we 507presented an imaging genetics methodological framework for 508 systematically identifying associations between genotypes and 509 imaging phenotypes, and demonstrated the utility of this method 510using the ADNI cohort. Our imaging genetics method can be broadly 511 summarized as the following four steps after quality control and 512preprocessing: (1) imaging phenotype definition, (2) GWAS of image 513phenotypes, (3) cluster and heat map analysis of imaging GWAS 514results, and (4) refined statistical modeling. 515

Imaging phenotype definition

Eight-six GM density ROI measures and 56 volume and cortical 517thickness ROI measures were extracted, using VBM and FreeSurfer 518methods respectively, and analyzed as image phenotypes in 519independent GWAS analyses. This approach is complementary to 520another recently proposed imaging genetics analysis method, voxel-521 wise GWAS (vGWAS) (Stein et al., submitted for publication). The 522vGWAS technique explores SNP associations with all voxels in the 523image space. Our study is ROI-based, analyzing fewer but anato-524mically meaningful imaging phenotypes and thus, requires less 525computational resources. In addition, we used multiple techniques 526to define imaging phenotypes. Among the top 5 SNPs identified 527as part of the present study (Table 4), rs10932886 (EPHA4), 528rs7610017 (TP63) and rs6463843 (NXPH1) are primarily associated 529 with VBM QTs, rs2075650 (TOMM40) is associated with FreeSurfer 530QTs, and rs429358 (APOE) is associated with ROIs extracted 531 using both techniques. These results suggest that the VBM and 532FreeSurfer QTs are not equally sensitive to the same genetic 533 markers and consequently may provide complementary informa-534tion. The VBM measures we employed are not modulated (Good 535 et al., 2001) and therefore measure GM densities (Ashburner and 536

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Fig. 2. Heat maps of SNP associations with quantitative traits (QTs) at the significance level of $p < 10^{-6}$. GWAS results at a statistical threshold of $p < 10^{-6}$ using QTs derived from FreeSurfer (top) and VBM/MarSBaR (bottom) are shown. $-\log_{10}(p$ -values) from each GWAS are color-mapped and displayed in the heat maps. Heat map blocks labeled with "x" reach the significance level of $p < 10^{-6}$. Only top SNPs and QTs are included in the heat maps, and so each row (SNP) and column (QT) has at least one "x" block. Dendrograms derived from hierarchical clustering are plotted for both SNPs and QTs. The color bar on the left side of the heat map codes the chromosome IDs for the corresponding SNPs. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

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Friston, 2000), which are different from the volume and thickness measures that FreeSurfer generates for analysis. The complementary nature of GM density, volumetric, and cortical thickness ROIs in assessing of early AD, MCI, and pre-MCI samples is consistent with our recent findings examining ADNI baseline MRI data (Risacher et al., 2009) as well as an independent cohort (Saykin et al., 2006).

GWAS of image phenotypes

Following quality control of the genotyping data, genome-wide 545 association studies were conducted on each of the 142 imaging 546 phenotypes. The entire set of the GWAS analyses was performed and 547 completed on a 112-node parallel computing environment within 20 548 min, suggesting an excellent potential for larger scale future 549 extensions. One extension could be to investigate more sophisticated 550

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Fig. 3. Manhattan and Q–Q plots of genome-wide association study (GWAS) of an example quantitative trait (QT). The QT examined in this analysis is the mean GM density of the right hippocampus (i.e., VBM phenotype RHippocampus, see Table 1) which was calculated using VBM/MarsBaR and adjusted for age, gender, education, handedness and ICV. Shown on the top panel is the Manhattan plot of the *p*-values ($-\log_{10}(observed$ *p*-value)) from GWAS analysis of the QT. The horizontal lines display the cutoffs for two significant levels: blue line for $p < 10^{-6}$, and red line for $p < 10^{-7}$. Shown on the bottom panel is the quantile–quantile (Q–Q) plot of the distribution of the observed *p*-values ($-\log_{10}(observed$ *p*-value)) in this sample versus the expected *p*-values ($-\log_{10}(expected$ *p*-value)) under the null hypothesis of no association. Genomic inflation factor (based on median chi-squared) is 1.01667. (For interpretation of the references to colour in this figure legend, the reader is referred to the web version of this article.)

statistical models (e.g., exploring SNP-by-SNP or SNP-by-diagnosis
 interactions). Another extension could be to involve more imaging
 phenotypes from other imaging modalities or longitudinal data.

554 Cluster and heat map analysis of imaging GWAS results

Heat maps and hierarchical clustering have been used frequently 555for grouping results in gene expression analysis for pattern discovery 556 (Eisen et al., 1998; Levenstien et al., 2003). In imaging genetics, heat 557maps can be equally useful for performing relevant pattern analysis 558 tasks thanks to the rich information contained within the maps and 559their effective mechanism to organize and visualize complicated 560imaging GWAS results. A straightforward use of a heat map is to select 561target OTs, SNPs, or associations for further analyses. Due to its 562563intuitive representation, some obvious patterns (e.g., the APOE SNP in Fig. 1) can be easily identified. For less obvious cases, other criteria 564could be used, for example, the selection of rs6463843 (NXPH1) 565because of its associations with multiple candidate phenotypic 566 regions (i.e., hippocampus) affected by AD (Fig. 2b). In addition, a 567 heat map can also be used to discover new patterns or structures. All 568 the QTs and SNPs are hierarchically clustered as dendrograms on the 569*x*-axis and *y*-axis, respectively. In the genomic domain, for those SNP 570 clusters that do not match the existing LD relationships, the 571 dendrogram provides the ability to identify novel inter-SNP structures 572(e.g., Sloan et al., submitted for publication). In the imaging domain, 573for those phenotype clusters that do not follow a regional or 574bilaterally symmetric pattern, there might be an opportunity to 575identify an underlying brain connectivity pattern associated with a 576genetic variation. 577

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(a) Comparison across all diagnostic groups, displayed at a threshold of p<0.01 (corrected with FDR)





Fig. 4. VBM genetics analysis for rs6463843 (NXPH1). A two-way ANOVA was performed on mean GM density maps to compare rs6463843 SNP genotype and baseline diagnostic group within the ADNI cohort. Analysis of the contrast of two genotype groups, GG>TT, is shown (n = 715; 166 AD (44 TT, 78 GT, 44 GG); 346 MCI (82 TT, 170 GT, 94 GG); 203 HC (35 TT, 105 GT, 63 GG)). Age, gender, education, handedness, and baseline ICV are included as covariates in all comparisons. Shown in the top panel (a) are the results of comparison involving all 715 subjects (i.e., across all the diagnostic groups), which are displayed at a threshold of p < 0.01 (corrected with FDR) with minimum cluster size (k) = 27. Shown in the bottom panel (b) are the results of comparisons within each of the three baseline diagnostic groups (AD, MCI, and HC), which are displayed at a threshold of p < 0.001 (uncorrected), with minimum cluster size (k) = 27.

578 Refined statistical modeling

In this paper, each heat map includes all the strong associations at 579a given significance threshold level, and can be used to guide further 580analyses using refined statistical models (e.g., involving diagnosis and 581other biomarkers, addressing interaction effects, etc.). These analyses 582can be performed using different strategies as follows: (1) select a 583target phenotype from the heat map and examine its whole genome 584mapping (e.g., Fig. 3); (2) pick a target SNP from a heat map and 585perform detailed image analysis (e.g., Fig. 4); and (3) choose a target 586SNP-QT association based on a heat map and/or an imaging analysis 587 588results, and perform a refined statistical modeling (e.g., Fig. 5). In this study, we conducted sample analyses for each of the above cases. The ultimate goal of these types of analyses is to identify genetic markers affecting brain structure and function, how these imaging and genetic markers interact with each other, as well as with diagnosis and/or other clinically and biologically relevant measures, and to gain a better understanding of disease risk and pathophysiology. 594

Imaging and genetics findings

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The APOE SNP rs429358 and TOMM40 SNP rs2075650 were 596 confirmed to be top markers affecting multiple brain structures in a 597

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Fig. 5. Refined analysis of sample imaging phenotypes in relation to rs6463843 (NXPH1) and baseline diagnosis. Two-way ANOVAs were applied to examine the effects of rs6463843 (NXPH1) and baseline diagnosis on four target GM density measures: (a–b) left and right hippocampal GMDs, and (c–d) left and right mean medial temporal lobe GMDs. All the analyses included age, gender, education, handedness and baseline ICV as covariates. *n* = 715 subjects were involved: 166 AD (44 TT, 78 GT, 44 GG); 346 MCI (82 TT, 170 GT, 94 GG); 203 HC (35 TT, 105 GT, 63 GG). The *p*-values for the main effect of diagnosis (DX), the main effect of SNP (SNP), and the interaction effect of SNP-by-diagnosis (DX × SNP) were shown in each plot.

mixed population of HC, MCI and AD (Farrer et al., 1997; Osherovich,
2009; Potkin et al., 2009a). Other SNPs, including rs10932886 (EPHA4),
rs7610017 (TP63) and rs6463843 (NXPH1), were also among the top
markers influencing brain structures in our analysis (Table 4). These
SNPs and the genes in which they are found or flank have a number of
important functions and potential pathways through which they may
influence the pathophysiological processes underlying AD.

605 The EPHA4 [EPH receptor A4] gene belongs to the ephrin receptor subfamily of the protein-tyrosine kinase family (Fox et al., 1995). The 606 interaction between neuronal EphA4 and glial ephrin-A3 was found to 607 bidirectionally control synapse morphology and glial glutamate 608 transport, which may ultimately regulate hippocampal function 609 610 (Carmona et al., 2009). In addition, EphA4 and EphB2 receptors 611 were reported to be reduced in the hippocampus before the development of impaired object recognition and spatial memory in 612 transgenic mouse models of AD (Simon et al., 2009). The TP63 [Tumor 613 protein 63] gene encodes a member of the p53 family of transcription 614 factors (Yang et al., 1998). A literature search did not locate any 615 articles associating TP63 with AD, cognitive impairment or neurode-616 generation. Additional imaging genetics analyses on both rs10932886 617 (EPHA4) and rs7610017 (TP63) appear warranted for future study. 618

619The NXPH1 [Neurexophilin 1] gene is a member of the neurex-
ophilin family and encodes a secreted protein which features a variable
N-terminal domain, a highly conserved, N-glycosylated central domain,
a short linker region, and a cysteine-rich C-terminal domain. This
protein forms a very tight complex with alpha neurexins, a group of
proteins that promote adhesion between dendrites and axons (Missler

and Sudhof, 1998). This gene has previously been implicated as a 625 candidate gene for neuroticism (van den Oord et al., 2008). In the 626 present study, a VBM analysis of rs6463843 (NXPH1) revealed 627 significantly reduced global and regional GM density in participants 628 with the TT genotype relative to those with the GG genotype. Additional 629 analyses indicated an interaction between rs6463843 (NXPH1) and 630 baseline diagnostic group in which AD patients homozygous for the T 631 allele were differentially vulnerable to decreased GM density in the 632 right hippocampus, a finding presumably reflecting greater atrophy 633 associated with this genotype in patients with AD. 634

Heat maps of imaging genetics associations at two significance 635 threshold levels $(p < 10^{-7} \text{ and } p < 10^{-6})$ were also reported. At the 636 conventional $p < 10^{-7}$ significance threshold, measures of hippocam-637 pal and amygdalar GM density and volume were strongly associated 638 with the APOE and TOMM40 SNPs. Ten additional imaging phenotypes 639 were strongly associated with at least one of the top SNPs (Fig. 1). We 640 also examined a somewhat less stringent threshold $(p < 10^{-6})$ in order 641 to identify additional SNP and imaging QT associations, as well as to 642 examine patterns of genotype and phenotype clustering. SNPs 643 associated with multiple unrelated or loosely related imaging 644 phenotypes may represent an interesting genetic marker affecting 645 overall brain structure or neurodegeneration. In addition, imaging 646 variables associated with a number of SNPs from multiple genes may 647 be particularly sensitive phenotypic markers for examining disease 648 associated genetic variation. Therefore, heat maps at multiple 649 statistical thresholds are useful in identifying candidate SNPs and 650 imaging phenotypes warranting further investigation. 651

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Limitations and future directions 652

The majority of analyses presented in this study focused on the 653 654 extraction and evaluation of imaging phenotypes and the relationship of genetic variation to these phenotypes. However, we also included a 655 limited assessment of the effects of baseline diagnostic group and the 656 interaction effect of SNP and diagnosis in the analysis of candidate 657 SNPs and phenotypes. Future studies could incorporate additional 658 659 variables (e.g., clinical measures, other types of imaging and biomarkers) in the GWAS design to examine their effects and 660 661 interactions with SNPs and/or target imaging phenotypes. The 662 present analysis did not address epistasis or gene-gene interactions, 663 a potentially very important topic. Future analyses should include 664 models that incorporate epistatic interactions which are likely to be important for understanding susceptibility and protective factors in 665 AD and other complex diseases. 666

Although we employed reasonably stringent thresholds for 667 assessing genome-wide significance, a large number of ROIs represent 668 a multiple comparison problem. The issue of determining the proper 669 statistical threshold for a whole genome and whole brain search for 670 associations is a challenging area for investigation (Nichols and 671 Holmes, 2002; Nichols and Inkster, 2009; Stein et al., submitted for 672 673 publication). The issue is complicated by the fact that variables within 674 both the genomic and neuroimaging dimensions are non-independent due to LD and spatial autocorrelation, respectively. The 675 determination of the effective number of independent statistical 676 tests under these conditions is an area of investigation. Models for the 677 678 joint distribution of both dimensions under the null hypothesis require development and validation. 679

Replication of current and future GWAS results in independent 680 samples will remain of critical importance for confirmation. Although 681 682 our follow-up analyses examine additional statistics at a more detailed level for yielding additional insights, these statistics are 683 non-independent of the statistics used to select candidate ROIs and 684 candidate associations. Given the recent interest in the non-685 independent analysis issue (e.g., Kriegeskorte et al., 2009), indepen-686 dent datasets for replication will be important for future studies to 687 688 confirm the findings. For the current ADNI sample, given its modest size, we were unable to use one half of the data for hypothesis 689 generation and the other half for confirmation, since one half of the 690 data (i.e., n = 367 in this study) cannot provide sufficient power to 691 692 detect moderate/small genetic effects (Potkin et al., 2009b). With additional replication and extension opportunities under develop-693 ment, we anticipate that there will be ample statistical power and the 694 ability to replicate potentially important findings in multiple 695 independent data sets in the future. 696

697 At present there are few opportunities for replication of imaging genetics results such as those emerging from ADNI given the unique 698 nature of this multi-dimensional data set. Fortunately, a worldwide 699 ADNI consortium is actively being developed and large scale 700 international data sets are likely to become available in the next few 701 702 years that can provide adequate replication samples. In addition, the 703 new NIH sponsored AD Genetics Consortium (ADGC) is assembling large meta-analytic databases of GWAS results that can provide 704confirmation of novel findings. Finally, the AlzGene meta-analytic 705 database (www.alzgene.org) of candidate genes for AD, curated by 706 707 Lars Bertram and colleagues (Bertram et al., 2007), provides a regularly updated source for determining the replication and 708 validation status of AD genes. 709

The AAL atlas (Tzourio-Mazoyer et al., 2002) used to create the 710 ROIs for the VBM analysis in this study is based on a single individual. 711 To take anatomical variability into account, an important future 712 direction will be to employ a probabilistic atlas, e.g., the Harvard-713 Oxford atlas (distributed with the FSL software package; http://fsl. 714 fmrib.ox.ac.uk/fsl/), or the LONI probabilistic brain atlas (Shattuck et 715 716 al., 2008). The most appropriate method to derive a GM-based summary statistic (e.g., density or volume) for a probabilistic ROI is a 717 topic warranting investigation. 718

Despite the limitations and challenges, the encouraging experi-719 mental results obtained using the proposed analytic framework 720 appear to have substantial potential for enabling the discovery of 721 imaging genetics associations and for localizing candidate imaging 722 and genomic regions for refined statistical modeling and further 723 characterization. Ultimately, imaging genetics holds the promise of 724 providing important clues to pathophysiology that could inform 725development of methods for earlier detection and therapeutic 726 intervention. 727

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